



## Unexpected Reactions of (*IR*)2,3,4,6-Tetra-O-acetyl-1-azido-D-galactopyranosyl Cyanide and the Derived Carboxamide with Triphenylphosphine

József Kovács<sup>a</sup>, István Pintér<sup>a\*</sup>, Mária Kajtár-Peredy<sup>a</sup>, László Somsák<sup>b</sup>

<sup>a</sup>Central Research Institute for Chemistry, Hungarian Academy of Sciences, H-1525 Budapest, P.O.Box 17, Hungary

<sup>b</sup>Department of Organic Chemistry, Lajos Kossuth University, H-4010 Debrecen, P.O.Box 20, Hungary

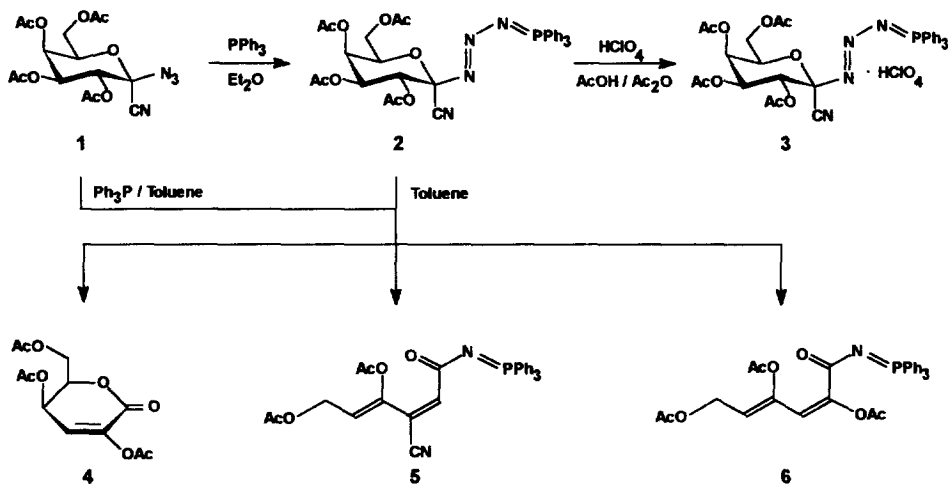
**Abstract:** Staudinger reaction of acetylated (*IR*)-1-azido-D-galactopyranosyl cyanide (**1**) with triphenylphosphine in diethyl ether led to the isolation of a crystalline phosphazide (**2**), unprecedented in carbohydrate chemistry. Spontaneous decomposition of **2** in toluene furnished the mixture of the unsaturated lactone **4**, as minor product, and two major products: the triphenylphosphoranylidene derivatives of (*2Z,4Z*)-3-cyano-4,6-diacetoxy-2,4-hexadienoic amide (**5**), and (*2E,4Z*)-triacetoxy-2,4-hexadienoic amide (**6**), respectively, owing to an unusual pyranoid ring opening between C-5 and the pyranose oxygen. The carboxamide analogue (**10**) of **1** underwent a regular phosphinimine formation affording the equilibrium mixture of both anomers **11** and **12**.

© 1997 Published by Elsevier Science Ltd.

The Staudinger reaction<sup>1</sup> - transformation of organic azides with tertiary phosphines to produce iminophosphoranes (phosphinimines) - is a versatile tool in organic syntheses.<sup>2</sup> The use of the phosphinimine method in the carbohydrate field provides an easy access to various N-containing sugars (carbodiimides, cyclic carbamates, epimines, ureido- and guanidino derivatives e.t.c.).<sup>3</sup> In the course of our studies on the synthesis and transformation of sugar phosphinimines, recently, a particular interest has been aimed at the Staudinger reaction of glycosyl azides bearing an additional functional group at the anomeric carbon. First, we described the reaction of tetra-O-acetyl-D-glucopyranosylidene 1,1-diazide<sup>4</sup> with triphenylphosphine to give a fused  $\nu$ -triazolo-pyranosyl phosphinimine.<sup>5</sup>

Now we report on the anomalous Staudinger reaction of (*IR*)2,3,4,6-tetra-O-acetyl-1-azido-D-galactopyranosyl cyanide<sup>6</sup> (**1**) and its carboxamide analogue<sup>6b</sup> **10**, respectively. Reaction of **1** with triphenylphosphine in molar ratio 1:1 in dry diethyl ether did not give the expected phosphinimine but the phosphazide **2** which precipitated from the reaction mixture in 80% yield. Phosphazides, as primary adducts of the Staudinger reaction, were isolated in several cases,<sup>7</sup> however, to our best knowledge **2** is the first one isolated in the sugar field. <sup>1</sup>H-NMR data of **2** exhibited signals of the peracetylated galactopyranose moiety (Table 1). The <sup>31</sup>P-NMR spectrum of **2** displayed a signal at  $\delta$  25.92 (Table 2) which is in good agreement with previously reported values of phosphazides.<sup>7,8</sup> Unlike sugar phosphinimines,<sup>3c,d,5</sup> the anomeric carbon of **2** ( $\delta$  94.27, Table 2) has no coupling with the phosphorus atom, as a consequence of the four bonds sequence between them.

The outcome of the reaction is practically not influenced by the molar ratio of the reactants; both with 2 and 0.5 equiv triphenylphosphine the yield of **2** (calculated for **1**) was 74% and 33%, respectively, while the



reactant used in excess was not consumed. Formation of **2** from **1** accords well with the anomalous Staudinger reaction of  $\alpha$ -azidodiphenylacetonitrile which furnished a phosphazide with *Z* configuration<sup>7</sup>. Similar geometry of the phosphazide moiety of **2** might be assumed, but in the lack of suitable crystals for X-ray analysis it could not be proved.

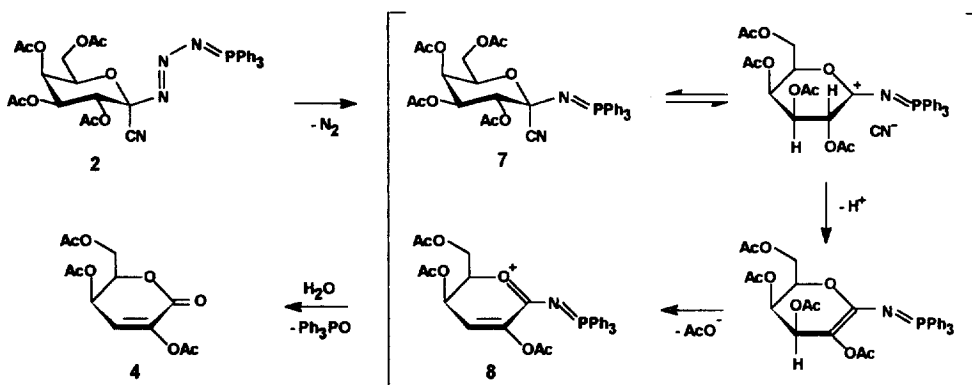
On treatment with perchloric acid in acetic anhydride **2** was transformed into the protonated salt **3**, providing evidence for the phosphazide structure of the molecule. Both **2** and its perchlorate salt (**3**) are stable in solid state, however in chloroform solution they decompose to give multicomponent mixtures.

Attempts to transform **2** into the corresponding phosphinimine by heating in toluene led to a dark complex mixture with tarry precipitation. However, when a solution of **2** in toluene was left to stand at room temperature for 3 d, tlc showed the disappearance of the phosphazide and formation of several products. The main components of the reaction mixture could be separated by column chromatography to give the unsaturated lactone **4**<sup>9,10</sup> (9%), the crystalline  $(2Z,4Z)$ -3-cyano-4,6-diacetoxy-*N*-(triphenylphosphoranylidene)-2,4-hexadienoic amide (**5**, 44 %) and  $(2E,4Z)$ -2,4,6-triacetoxy-*N*-(triphenylphosphoranylidene)-2,4-hexadienoic amide (**6**, 18%), besides triphenylphosphine oxide (15%).

$^1\text{H}$ -,  $^{13}\text{C}$ - and  $^{31}\text{P}$ -NMR spectra of the unsaturated aliphatic phosphinimines **5** and **6** revealed the constitution of both molecules and pointed to the close analogy of their structure (Tables 1 and 2). The exact structure of **5** was proved by X-ray diffraction.<sup>12</sup> In the case of **6** *Z* configuration of the C-4 + C-5 double bond was corroborated by NOE measurement, indicating the closeness of H-3 and H-5. Otherwise, the large value of the coupling  $^3J_{\text{CO,H-3}} = 7.8$  Hz, determined by selective 2D INEPT measurement<sup>13</sup>, proved the *trans* relationship of H-3 and the imide carbonyl, which corresponds to the *2E* configuration. Thus, disregarding the CN-substituent in **5**, the iminophosphoranes **5** and **6** contain the same carbon skeleton.

When the reaction of **1** with triphenylphosphine (molar ratio 1:1.05) was performed in dry toluene the same products were obtained as from the isolated phosphazide, but in this case the triacetate **6** was formed predominantly. The hexadienoic phosphinimines **5** and **6** are quite stable compounds. In contrast to sugar phosphinimines they do not react with carbon dioxide, owing to their resonance-stabilized structure.

As for the reaction mechanism, transformation of the phosphazide (**2**) in solution may proceed in two different ways. The side-reaction (Scheme 1) leading to the unsaturated lactone **4** involves the formation of 1-cyano-1-phosphinimine **7** which can be stabilized by releasing the cyanide anion and eliminating acetic acid to give an unsaturated lactone-imino phosphonium salt (**8**). Formation of the enollactone **4** is attributed to the hydrolysis of **8** during the reaction or chromatographic separation.

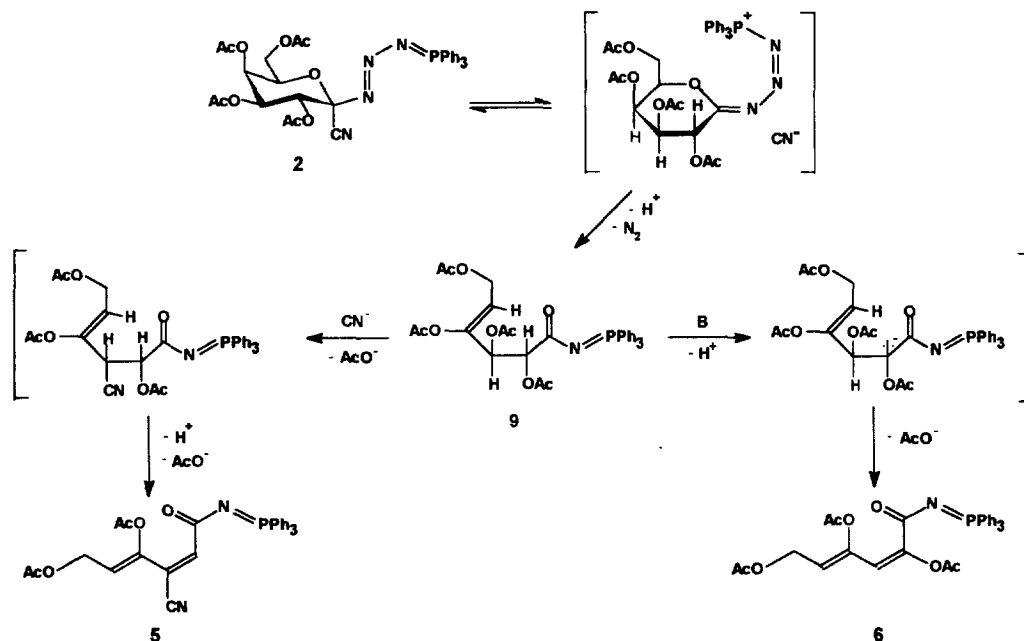


Scheme 1

The main pathway (Scheme 2) resulting in the formation of acyclic unsaturated iminophosphoranes **5** and **6** requires an unusual ring opening between C-5 and the pyranose oxygen. The reason for this might be the relative stability of the phosphazide system in the *Z* configuration which allows an interaction between the pyranose oxygen and the positively charged phosphorus atom. This anchimeric effect may help on the pyranose ring opening. Subsequent deprotonation at C-4 and splitting of nitrogen make the reaction irreversible furnishing a 4,5-unsaturated aldonyl phosphinimine (**9**) which may serve as a key intermediate for both final products. On one hand, replacement of AcO-3 by the cyanide anion ( $S_N2$  reaction) followed by release of H-3 and subsequent  $\beta$ -elimination of the acetate anion from C-2 leads to the product **5**. Alternatively, **9** may be stabilized by deprotonation from C-2 and  $\beta$ -elimination of AcO-3 to give the triacetate **6**. The influence of the reaction conditions on the yields of the products is being investigated.

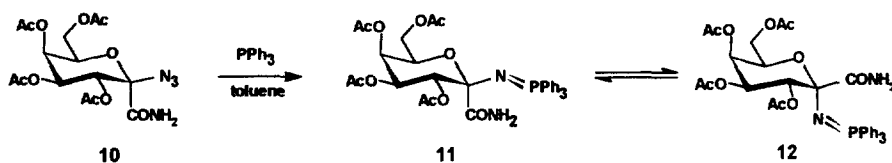
A similar acetoxy-group elimination process affording heterocycles with penta-dienyl side chain was observed<sup>14</sup> in the anomalous Wittig reaction of aldonic thioamide derivatives.

In contrast to the very fast transformation of **1** with triphenylphosphine, the carboxamide analogue **10**<sup>6b</sup> reacts very slowly under the same conditions. With 1.1 equiv triphenylphosphine in diethyl ether at room



Scheme 2

temperature the reaction was not complete even in two weeks. Therefore, 2 equiv triphenylphosphine was used in toluene at 80° C for 10 h and the reaction mixture was chromatographed to give 3,4,5,7-tetra-O-acetyl-2-deoxy-2-(triphenylphosphoranylideneamino)-β-D-galacto-hept-2-ulopyranosonamide (**11**) and its α-anomer (**12**) in 36% and 21% yield, respectively. Both anomers are stable in neat form, however, during six weeks in chloroform solution they anomerize to give a 15:85 mixture of **11** and **12** as shown by NMR measurements. The preponderance of **12** in the equilibrium mixture - indicated also by tlc and the change of optical rotation - may be explained by the strong anomeric effect of the phosphinimino-group<sup>15</sup> and the reverse anomeric effect of the carbamoyl group.<sup>16</sup>



The <sup>1</sup>H-NMR spectra (Table 1) of both **11** and **12** indicated the acetylated galactopyranose moieties to be in the <sup>4</sup>C<sub>1</sub> conformation. The theoretically possible ring closure between the phosphinimine function and the carbamoyl group to form a spirobicycle could be ruled out by the heterocorrelated 2D <sup>1</sup>H-<sup>15</sup>N NMR spectrum of the equilibrium mixture of **11** and **12** which proved that in both anomers the two NH protons belong to the same nitrogen atom. Selective <sup>1</sup>H-<sup>1</sup>H NOE measurements helped to assign the stereochemistry of **11** and **12**.

Table 1. <sup>1</sup>H-NMR data<sup>a</sup> measured for CDCl<sub>3</sub> solutions at 400 MHz (δ [ppm], J [Hz])

	H-2 (J <sub>2,3</sub> )	H-3 (J <sub>3,4</sub> )	H-4 (J <sub>4,5</sub> )	H-5 (J <sub>5,6a</sub> )	H-6a (J <sub>6a,6b</sub> )	H-6b (J <sub>5,6b</sub> )	Phenyl H <sub>ortho</sub> H <sub>para</sub> H <sub>meta</sub>	Acetyl	NH (J <sub>NH,NH1</sub> )
<b>2</b>	5.73 (10.5)	5.29 (3.2)	5.52 (1.3)	4.42 (6.8)	4.20 (11.2)	4.13 (6.2)	7.72 7.64 7.52	2.14, 2.02, 1.97, 1.67	--
<b>3</b>	5.47 (10.5)	5.22 (3.2)	5.51 (1.3)	4.38 (6.3)	4.15 (11.3)	4.12 (6.5)	7.7-7.8 7.89 7.7-7.8	2.17, 2.04, 1.97, 1.81	--
<b>4</b>	--	6.64 (6.3)	5.43 (2.6)	4.92 (6.0)	4.37 (11.6)	4.35 (6.5)	--	2.28, 2.12, 2.11	--
<b>5</b>	6.91	--	--	6.05 (6.5)	4.51	4.51 (6.5)	7.74 7.61 7.50	2.01, 1.61	--
<b>6</b>	--	6.10 <sup>b</sup>	--	5.64 (6.9)	4.53	4.53 (6.9)	7.76 7.56 7.47	2.21, 1.97, 1.71	--
<b>11</b>	5.12 (10.7)	5.95 (3.6)	5.38 (2.0)	5.03 (6.4)	3.69 (10.9)	3.66 (6.8)	7.69 7.49 7.43	2.11, 1.90, 1.88, 1.84	8.03, 5.70 (6.0)
<b>12</b>	5.41 <sup>c</sup> (10.2)	5.83 (3.5)	5.59 (1.5)	4.78 (5.6)	4.12 (11.2)	4.00 (7.3)	7.76 7.46 7.39	2.12, 2.00, 1.98, 1.91	6.20, 4.67 (4.0)

<sup>a</sup>Listed according to parent sugar numbering. <sup>b</sup>J<sub>3,5</sub> ≈ 1 Hz. J<sub>5,6</sub> ≈ 1 Hz. <sup>c</sup>J<sub>H,2,P</sub> = 5.0 Hz.

Table 2. <sup>13</sup>C- and <sup>31</sup>P-NMR data<sup>a,b,c</sup> for CDCl<sub>3</sub> solutions<sup>d</sup>

	C-1	C-2	C-3	C-4	C-5	C-6	C <sub>ipso</sub>	C <sub>ortho</sub>	C <sub>meta</sub>	C <sub>para</sub>	CH <sub>2</sub> CO	CH <sub>3</sub> CO	CN	CONH <sub>2</sub>	P
<b>2</b>	94.27	68.21	70.28	66.80	71.55	61.05	125.90 (95.9)	133.67 (9.2)	128.98 (12.0)	133.10 (2.9)	170.26 170.20 169.90 168.26	20.67 20.62 20.57 20.30	114.49	--	25.92
<b>3</b>	92.85	66.13 <sup>e</sup>	69.47	66.79 <sup>e</sup>	72.68	60.58	117.06 (102)	134.34 (11.7)	130.40 (13.9)	136.23 (3.0)	170.19 169.92 169.61 168.25	20.62 20.60 20.44 20.16	112.13	--	39.00
<b>4</b>	157.53	141.85	124.42	62.97	76.13	61.27	--	--	--	--	170.39 169.76 168.08	20.67 20.51 20.35	--	--	--
<b>5</b>	172.10 (7.4)	144.19 (23.6)	111.17 (2.6)	142.41	119.25	58.65	126.96 (99.5)	133.21 (10.3)	128.88 (12.5)	132.77 (2.9)	170.53 167.72	20.75 19.92	117.07	--	20.91
<b>6</b>	169.88 (7.0)	148.15 (25.3)	116.82 (1.4)	144.47	117.35	58.99	127.48 (99.7)	133.21 (10.1)	128.68 (12.5)	132.41 (2.7)	170.71 169.34 168.26	20.84 20.81 20.19	--	--	22.08
<b>11</b>	90.65 (8.7)	74.13 (2.6)	69.49	68.31	70.80	62.02	132.70 (101.8)	132.84 (10.1)	128.10 (12.2)	131.27 (2.7)	170.49 170.27 169.97 169.92	21.16 20.90 20.75 20.73	--	174.52	5.20
<b>12</b>	91.15 (1.9)	71.01 (16.8)	69.92	69.22	67.14	62.73	132.70 (102.1)	132.81 (9.8)	128.00 (12.1)	131.09 (2.7)	170.54 170.47 170.44 170.39	20.88 20.87 20.81 20.67	--	172.58	8.68

<sup>a</sup>Listed according to parent sugar numbering. <sup>b</sup>Chemical shifts (δ [ppm]). <sup>c</sup>Couplings [Hz] of <sup>31</sup>P with the corresponding <sup>13</sup>C in parenthesis. <sup>d</sup>Recorded at 101 MHz (<sup>13</sup>C) and 162 MHz (<sup>31</sup>P), respectively. <sup>e</sup>Assignments may be interchanged.

In **11** NOE-s were measured between H-2 and the *ortho* phenyl protons, while irradiation of H-3 resonance gave enhancement of the amide proton signals. On the contrary, spatial proximity was observed between H-2 and the amide protons, moreover, between H-3, H-5 and the phenyl protons in **12**. These results are in agreement with the vicinal coupling value between the amide carbonyl and H-2 which, knowing the conformation ( ${}^4C_1$ ) of the pyranoid ring, allowed to establish the absolute configuration of the anomeric carbon in both anomers. Thus,  ${}^3J_{\text{CONH}_2, \text{H}-2} = 5.7$  Hz measured in **11** corresponds to the antiperiplanar (*trans*) orientation of H-2 and the amide carbonyl (*S* configuration), while  ${}^3J_{\text{CONH}_2, \text{H}-2} = 1.5$  Hz for **12** indicates a *gauche* coupling (*R* configuration). These values accord well with the corresponding three bond couplings of the parent azido-amide **10** and those of other glycopyranosylidene derivatives having a C-substituent at the anomeric carbon.<sup>6</sup> Both C-1 and C-2 (according to parent sugar numbering) couple with phosphorus, however, a striking difference was found in the  ${}^2J_{\text{C}-1, \text{P}}$  value for **11** (8.7 Hz) and that for **12** (1.9 Hz) and, especially, in the  ${}^3J_{\text{C}-2, \text{P}}$  value for **11** (2.6 Hz) and that for **12** (16.8 Hz). On the basis of the stereospecificity of  ${}^3J_{\text{C}, \text{P}}$  couplings<sup>17</sup> the latter large value indicates the  $\gamma$ -*anti* orientation of P-N and C-1 + C-2 bonds in the molecule **12**, stabilized by the *exo*-anomeric effect<sup>18</sup> In contrast, the small value of  ${}^3J_{\text{C}-2, \text{P}}$  in **11** reflects a conformation in which the orientation of the P-N bond is  $\gamma$ -*gauche* related to the C-1 + C-2 bond. The latter conformation being opposite to that measured in acetylated glycosyl phosphinimines<sup>3c,d</sup> might be stabilized by an interaction between phosphorus and the oxygen atom of the carbamoyl group. Of course, this stabilizing P-O interaction can also work in the case of the thermodynamically more stable anomer **12**. The  ${}^{31}\text{P}$ -NMR spectra of **11** and **12** exhibited signals of phosphorus at very high field ( $\delta$  5.20 in **11** and  $\delta$  8.60 in **12**) in comparison to those of various imino-phosphoranes.<sup>3c,d,7</sup> This may be the consequence of the special structural feature that in **11** and **12** the phosphinimino group is bonded to an electronegative quaternary carbon.

These sugar phosphinimines of new type have low reactivity, both **11** and **12** remained intact on treatment with methyl iodide as well as in the attempted aza-Wittig reaction with carbon dioxide. This latter reaction was unsuccessful, as reported very recently<sup>19</sup>, even if the azido-amide **10** was treated with tri-*n*-butyl-phosphine in the presence of carbon dioxide. On the contrary, the phosphinimine method has been successfully applied to a *D-ribo* configured, furanoid azido-amide providing a facile synthetic route to (+)hydantocidin<sup>20</sup>

## EXPERIMENTAL

Tlc was performed on DC-Alurolle, Kieselgel 60 F<sub>254</sub> (Merck); detection by UV light and charring with H<sub>2</sub>SO<sub>4</sub>. For column chromatography Kieselgel 60 (Merck) was used. Melting points were measured in open capillary tubes in a Büchi apparatus or on a Koffler hot-stage and are uncorrected. Optical rotations were determined with a Zeiss Polamat A polarimeter at 25 °C. Ir spectra were taken with a Nicolet FT 205 spectrometer. The Raman spectra were recorded on a Nicolet 950 FT Raman spectrometer. NMR spectra were recorded on a Varian VXR-400 spectrometer. Chemical shifts refer to signals of tetramethylsilane in the case of  ${}^1\text{H}$ - and  ${}^{13}\text{C}$  spectra and to 85 % aqueous phosphoric acid in the case of  ${}^{31}\text{P}$  spectra. Fast-atom bombardment (FAB) mass spectra were obtained with a VG ZAB-2SEQ mass spectrometer (using 3-nitro-benzylalcohol matrix). Microanalyses were performed in the Microanalytical Laboratory of the Institute

(1R)2,3,4,6-Tetra-O-acetyl-1-(3-triphenylphosphazido)-D-galactopyranosyl cyanide [3,4,5,7-tetra-O-acetyl-2-deoxy-2-(3-triphenylphosphazido)-β-D-galacto-hept-2-ulopyranosono-nitrile] **2**. (a) To a stirred solution of **1**<sup>6</sup> (159 mg, 0.4 mmol) in dry diethyl ether (3 ml) was added a solution of triphenylphosphine (110 mg, 0.42 mmol) in the same solvent (4 ml). A white solid precipitated within 1 min. After stirring for 10 min at room temperature the product was collected and washed with dry ether to give practically pure **2** (211 mg, 80 %), mp 92-93 °C;  $[\alpha]_D^{+99}$  (c1.5, CHCl<sub>3</sub>); IR (KBr):  $\nu$  1756 (OAc), 1430, 1113, 723, 694 cm<sup>-1</sup> (Ph), Raman (solid):  $\nu$  2235 (CN), 1756 (OAc), 1586, 1103, 1027, 997 cm<sup>-1</sup> (Ph); MS: 661 [M+H]<sup>+</sup>. Anal. Calcd for C<sub>33</sub>H<sub>33</sub>N<sub>4</sub>O<sub>9</sub>P (660.64): C, 60.00; H, 5.04; N, 8.48. Found: C, 59.83; H, 4.89; N, 8.13.

(b) When the reaction was carried out in the same way but using 2 equiv triphenylphosphine, **2** was obtained in 74 % yield, identical with the product in (a). Excess of PPh<sub>3</sub> was detected (tlc) in the mother liquor.

(c) Using 0.5 equiv PPh<sub>3</sub> in the same reaction afforded **2** in 33 % yield (calculated for **1**), identical with the product in (a). Concentration of the filtrate and crystallization of the residue from EtOH gave unreacted **1** (37 %).

*Perchlorate salt (3) of phosphazide 2*. To a solution of **2** (0.33 g, 0.5 mmol) in an ice-cold mixture of aqueous 70 % perchloric acid (0.2 ml) and acetic anhydride (2 ml) was added diethyl ether (25 ml) to give crude **3** (0.34 g, 89 %), mp 129-133 °C. Precipitation with diethyl ether from dichloromethane solution afforded the pure salt (0.25 g, 66 %) mp 138 °C;  $[\alpha]_D^{+44}$  (c2, CHCl<sub>3</sub>); IR (KBr):  $\nu$  1761 (OAc), 1439, 1119, 732 (Ph), 1100, 623 cm<sup>-1</sup> (ClO<sub>4</sub><sup>-</sup>). Anal. Calcd for C<sub>33</sub>H<sub>34</sub>ClN<sub>4</sub>O<sub>13</sub>P (761.10): C, 52.08; H, 4.50; N, 7.36. Found: C, 51.94; H, 4.33; N, 7.26.

*Transformation of 2 in toluene*. (a) A solution of **2** (0.4 g, 0.6 mmol) in dry toluene (8 ml) was stored at room temperature for 3 d. The brown solution was decanted from the tarry residue and concentrated in *vacuo* to give a complex mixture. Column chromatography on silica gel (eluent: dichloromethane-acetone 9:1) afforded (5R,6R)-3,5-bis(acetoxy)-6-(acetoxymethyl)-5,6-dihydro-2H-pyran-2-one<sup>9,10</sup> **4** (15 mg, 9 %) as a colourless oil, the crystalline (2Z,4Z)-3-cyano-4,6-diacetoxy-N-(triphenylphosphoranylidene)-2,4-hexadienoic amide **5** (136 mg, 44 %) and (2E,4Z)-2,4,6-triacetoxy-N-(triphenylphosphoranylidene)-2,4-hexadienoic amide **6** (60 mg, 18 %) as a colourless solid and triphenylphosphine oxide (25 mg, 15 %), respectively.

For **4**:  $[\alpha]_D^{-150}$  (c0.5, CHCl<sub>3</sub>). Anal. Calcd for C<sub>12</sub>H<sub>14</sub>O<sub>8</sub> (286.24): C, 50.35; H, 4.93. Found: C, 50.58; H, 4.99.

For **5**: mp 137 °C (from diethyl ether); IR (KBr):  $\nu$  2227 (CN), 1769, 1741 (OAc), 1593 (CO), 1440, 1114, 723 cm<sup>-1</sup> (Ph); Raman (solid):  $\nu$  2232 (CN), 1671, 1609 1590 (C=C), 1027, 999 cm<sup>-1</sup> (Ph); MS: 513 [M+H]<sup>+</sup>. Anal. Calcd for C<sub>29</sub>H<sub>25</sub>N<sub>2</sub>O<sub>5</sub>P (512.51): C, 67.96; H, 4.92; N, 5.47. Found: C, 67.90; H, 5.03; N, 5.60.

For **6**: mp 128-129 °C (from diethyl ether); IR (KBr):  $\nu$  1763, 1737 (OAc), 1598 (CO), 1439, 1116, 723 cm<sup>-1</sup> (Ph); Raman (solid):  $\nu$  1656, 1590 (C=C), 1029, 1000 cm<sup>-1</sup> (Ph); MS: 546 [M+H]<sup>+</sup>. Anal. Calcd for C<sub>30</sub>H<sub>28</sub>NO<sub>7</sub>P (545.54): C, 66.05; H, 5.17; N, 2.57. Found: C, 66.09; H, 5.26; N, 2.74.

(b) To a solution of **1** (219 mg, 0.55 mmol) in dry toluene (5 ml) was added dropwise a solution of triphenylphosphine (152 mg, 0.58 mmol) in the same solvent (5 ml) during 15 min and the mixture was stored at room temperature for 3 d. Work-up according to (a) furnished **4** (9 mg, 6 %) and **5** (54 mg, 19 %) and **6** (88 mg, 29 %) and triphenylphosphine oxide (32 mg, 21 %), respectively, identical with the products in (a).

*3,4,5,7-Tetra-O-acetyl-2-deoxy-2-(triphenylphosphoranylideneamino)-β- (11) and -α-D-galacto-hept-2-ulopyranosonamide (12)*. To a solution of **10** (416 mg, 1 mmol) in dry toluene (25 ml) was added triphenylphosphine (0.52 g, 2 mmol) and the mixture was heated at 80 °C for 10 h. After removal of the solvent in *vacuo*, the residue was chromatographed on a silica gel column using diethyl ether as eluent. First, unreacted triphenylphosphine (239 mg, 46 %) was recovered then the anomeric mixture of **11** and **12** (483 mg, 75 %) was obtained. Repeated column chromatography of the raw product using dichloromethane/acetone 3:1 as eluent gave the syrupy **11** (237 mg, 36 %) and **12** (134 mg, 21 %), respectively.

For **11**:  $[\alpha]_D^{-12.9} \rightarrow +79.4$  (c3.1, CHCl<sub>3</sub>); IR (film):  $\nu$  1749 (OAc), 1685 (CONH<sub>2</sub>), 1438, 1109, 715, 696 cm<sup>-1</sup> (Ph). Anal. Calcd for C<sub>23</sub>H<sub>35</sub>N<sub>2</sub>O<sub>10</sub>P (650.63): C, 60.92; H, 5.42; N, 4.31. Found: C, 60.59; H, 5.27; N, 4.19.

For **12**:  $[\alpha]_D^{+95} \rightarrow +79.8$  (c3.2, CHCl<sub>3</sub>); IR(film):  $\nu$  1749 (OAc), 1700 (CONH<sub>2</sub>), 1438, 1110, 716, 695 cm<sup>-1</sup> (Ph). Anal. Calcd for C<sub>33</sub>H<sub>35</sub>N<sub>2</sub>O<sub>10</sub>P (650.63): C, 60.92; H, 5.42; N, 4.31. Found: C, 60.74; H, 5.19; N, 4.21.

Solutions of both **11** and **12** in CDCl<sub>3</sub> were proved by NMR to transform into anomeric mixtures of **11** and **12** with the equilibrium ratio of 15:85 within 6 weeks, in accord with the change of the optical rotations.

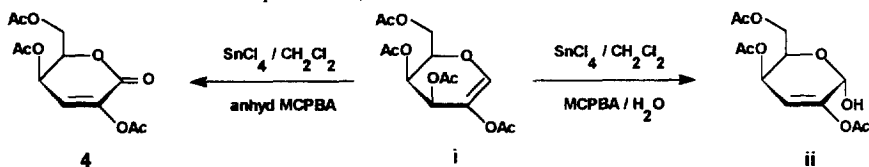


## ACKNOWLEDGEMENT

This work was supported by the Hungarian Scientific Research Fund (OTKA T 14939 and T 19339). We thank Dr. S. Holly for performing and discussing IR and Raman spectra, Mrs. Á. Gömörly for recording MS and Dr. L. Radics for heterocorrelated  $^1\text{H}$ - $^{15}\text{N}$  NMR measurements. The authors acknowledge Professor F. W. Lichtenthaler for the valuable discussions on the problem of the enollactone **4**.

## REFERENCES AND NOTES

1. Staudinger, H.; Meyer, J. *Helv. Chim. Acta* **1919**, *2*, 635-646.
2. a) Gololobov, Yu. G. *Tetrahedron* **1992**, *48*, 1353-1406; b) Molina, P.; Vilaplana, M. J. *Synthesis* **1994**, 1197-1218.
3. a) Messmer, A.; Pintér, I.; Szegő, F. *Angew. Chem.* **1964**, *76*, 227-228; b) Pintér, I.; Kovács, J.; Messmer, A.; Kálmán, A.; Tóth, G.; Lindberg, B. K. *Carbohydr. Res.* **1979**, *72*, 289-296; c) Kovács, J.; Pintér, I.; Messmer, A.; Tóth, G.; *Carbohydr. Res.* **1985**, *141*, 57-65; d) Kovács, J.; Pintér, I.; Messmer, A.; Tóth, G.; Duddeck, H. *Carbohydr. Res.* **1987**, *166*, 101-111; e) Cano, F. H.; Foces-Foces, C.; Jiménez-Barbero, J.; Allemany, A.; Bernabé, M.; Martín-Lomas, M. J. *Org. Chem.* **1987**, *52*, 3367-3372; f) Rück, K.; Kunz, H. *Synthesis* **1993**, 1018-1028; g) Pintér, I.; Kovács, J.; Tóth, G. *Carbohydr. Res.* **1995**, *273*, 99-108; h) Mészáros, P.; Kovács, J.; Pintér, I. *Carbohydr. Lett.* (in press).
4. a) Praly, J-P.; El Kharraf, Z.; Descotes, G. *J. Chem. Soc., Chem. Commun.* **1990**, 431-432; b) Praly, J-P.; Péquery, F.; Di Stéfano, C.; Descotes, G. *Synthesis* **1996**, 577-579.
5. Kovács, J.; Pintér, I.; Kajtár-Peredy, M.; Praly, J-P.; Descotes, G. *Carbohydr. Res.* **1995**, *279*, C1-C3.
6. a) Praly, J-P.; Di Stéfano, C.; Descotes, G.; Faure, R.; Somsák, L.; Eperjesi, I. *Tetrahedron Lett.* **1995**, *36*, 3329-3332; b) Somsák, L.; Sós, E.; Györgydeák, Z.; Praly, J-P.; Descotes, G. *Tetrahedron* **1996**, *52*, 9121-9136, and refs. therein.
7. Molina, P.; López-Leonardo, C.; Llamas-Botía, J.; Foces-Foces, C.; Fernández-Castano, C. *Tetrahedron* **1996**, *52*, 9629-9642, and refs. therein.
8. Ponomarchuk, M. P.; Kasukhin, L. F.; Shevchenko, M. V.; Sologub, L. S.; Kukhar, V. P. *Zh. Obshch. Khim.* **1984**, *54*, 2468-2473.
9. Lichtenthaler, F. W.; Rönninger, S.; Jarglis, P. *Liebigs Ann. Chem.* **1989**, 1153-1161.
10. The physical and spectral data for the enollactone **4** are at variance with those reported for a product of structure **4** with mp 159-161 °C and  $[\alpha]_D$  -45 (c1,  $\text{CHCl}_3$ ), obtained on  $\text{SnCl}_4$  catalyzed 3-chloroperbenzoic acid (MCPBA)-oxidation of 3,4,6-tri-O-acetyl-2-deoxy-D-galactal<sup>9</sup> (i). These discrepancies were resolved by the finding<sup>11</sup> that  $\text{SnCl}_4$ -promoted peroxidation of (i) with anhydrous MCPBA indeed yields the enollactone **4** as colourless syrup of  $[\alpha]_D$  -151.5 (c1,  $\text{CHCl}_3$ ), exhibiting the NMR data reported here (Tables 1 and 2). The crystalline product previously obtained and erroneously taken for enollactone **4**, proved to be<sup>11</sup> 2,4,6-tri-O-acetyl-3-deoxy- $\alpha$ -D-threo-hex-2-enopyranose (ii) of mp 160-161 °C and  $[\alpha]_D$  -45 (c1,  $\text{CHCl}_3$ );  $^1\text{H-NMR}$  ( $\text{CDCl}_3$ ):  $\delta$  2.07, 2.09, 2.19 (3s, OAc), 4.17 (m, H-5), 4.32 (m, H<sub>2</sub>-6), 5.26 (dd, H-4), 5.50 (s, H-1), 6.02 (d, H-3),  $J_{3,4}$  = 6.1,  $J_{4,5}$  = 1.3 Hz.<sup>11</sup> Compound ii is formed instead of **4**, when the  $\text{SnCl}_4$ -induced MCPBA-oxidation is not performed under strictly anhydrous conditions (commercial MCPBA contains water up to 35 %).



11. Lichtenthaler, F. W. personal communication.
12. Argay, Gy.; Kálmán, A.; Kovács, J.; Pintér, I. in preparation.
13. Jippo, T.; Kamo, O.; Nagayama, K. *J. Magn. Reson.* **1986**, *66*, 344-348.

14. Somogyi, L.; Herczegh, P.; Batta, Gy. *Heterocycles* **1984**, *24*, 2735-2738.
15. Paulsen, H.; Györgydeák, Z.; Friedmann, M. *Chem. Ber.* **1974**, *107*, 1590-1613.
16. Chmielewski, M.; BeMiller, J. N.; Cerretti, D. P. *J. Org. Chem.* **1981**, *46*, 3903-3908.
17. a) Buchanan, G. W.; Morin, F. G. *Can. J. Chem.* **1979**, *57*, 21-26; b) Quin, L. D.; Gallagher, M. J.; Cunkle, G. T.; Chesnut, D. B. *J. Am. Chem. Soc.* **1980**, *102*, 3136-3143; c) Duncan, M.; Gallagher, M.J.; *Org. Magn. Reson.* **1981**, *15*, 37-42.
18. Juaristi, E.; Cuevas, G. *Tetrahedron* **1992**, *48*, 5019-5087, and refs therein.
19. Ősz, E.; Sós, E.; Somsák, L.; Szilágyi, L.; Dinya, Z. *Tetrahedron* **1997**, *53*, 5813-5824.
20. Mio, S.; Kumagawa, Y.; Sugai, S. *Tetrahedron* **1991**, *47*, 2133-2144.

(Received in UK 4 August 1997; revised 26 August 1997; accepted 28 August 1997)